

A highly efficient synthesis of unnatural L-sugars from D-ribose

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Abstract—A preparative and short synthesis of L-ribose and L-apiose was accomplished starting from D-ribose via stereoselective *cis*-dihydroxylation and C2-hydroxymethylation, respectively. These L-sugars can serve as versatile intermediates for the synthesis of L-nucleosides.

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1. Introduction

Since the discovery of L-β-1,3-oxathiolanyl cytosine (Lamivudine, 3TC)¹ as anti-hepatitis B virus (HBV) drug, L-nucleosides have opened a new era for the development of new antiviral agents. As a result, L-FMAU has been found to be active against HBV and is now undergoing clinical trials.² L-2',3'-Dideoxy-2',3'-dideoxy-5-fluorocytidine (L-Fd4C)³ exhibited potent anti-HBV activity and is also in clinical trials. L-2',3'-Dideoxycytidine (L-ddC) showed potent anti-HBV activity and its 5-fluorocytosine derivative exhibited potent antiviral activity against HIV-1 and HBV.⁴ Recently, L-thymidine (L-dT) and L-2'-deoxycytidine (L-dC) were found to be active against HBV (Fig. 1).^{5,6} The main advantage of these L-nucleosides is to show much less cytotoxicity than their counterparts, D-nucleosides, while maintaining antiviral activity.

Although all these nucleosides may be synthesized from L-ribose, L-ribose may not be used as a starting material because of high price, indicating that an efficient synthesis of L-ribose is highly desirable. Although several good synthetic routes to L-ribose have been reported from D-carbohydrates such as D-ribose,⁷ D-glucose,⁸ D-galactose,⁹ and D-mannose,¹⁰ we wanted to develop an alternative

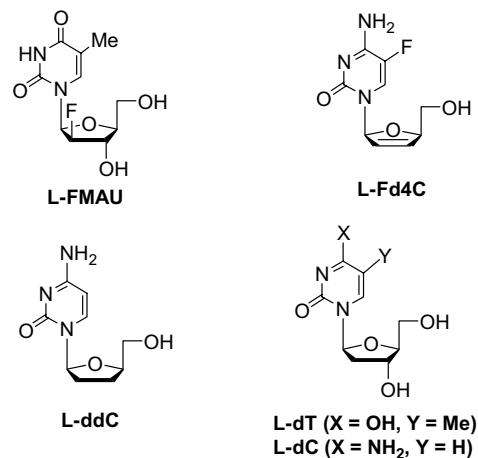


Figure 1. Antiviral active L-nucleosides.

and practical synthetic methodology to L-ribose, which can be widely utilized for the synthesis of biologically active compounds including L-nucleosides.

On the other hand, L-apiose is also unnatural sugar which has been used for the development of biologically active materials.¹¹ However, L-apiose is also too expensive to be used for a starting material for the development of biologically active compounds. Thus, it was also of great interest to develop a preparative synthetic method of L-apiose since only a few synthetic methods¹² have been reported so far. Herein, we wish to describe

Keywords: L-Ribose; L-Apiose; *cis*-Dihydroxylation; Hydroxymethylation.

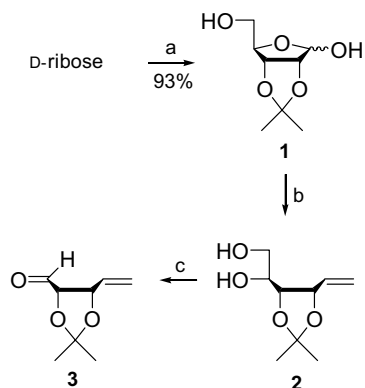
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an efficient and practical synthesis of L-sugars from D-ribose.

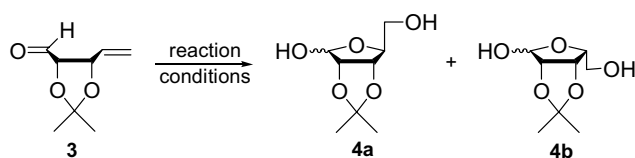
2. Results and discussion

As shown in Scheme 1, the preparative synthesis of L-ribose started from D-ribose. D-Ribose was converted to its 2,3-acetonide **1**¹³ by the conventional method. Treatment of **1** with methyltriphenylphosphonium bromide in the presence of potassium *t*-butoxide afforded vinyl diol **2**. Oxidative cleavage of **2** with sodium metaperiodate produced highly volatile vinyl aldehyde **3**,¹³ which was a good substrate for *cis*-dihydroxylation.

As shown in Scheme 2, several dihydroxylation reactions on vinyl aldehyde **3** were attempted to achieve

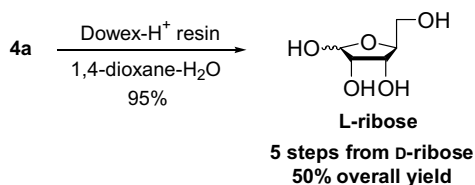


Scheme 1. Synthesis of the key intermediate. Reagents and conditions: (a) acetone, *c*-H₂SO₄, rt, 2.5 h; (b) Ph₃PCH₃Br, K-*t*-OBu, THF, rt, 15 h; (c) NaIO₄, H₂O, CH₂Cl₂, rt, 15 min.



reaction conditions	(4a : 4b) ^a
OsO ₄ , NMO, H ₂ O	6.67 : 1
AD-mix α	6.25 : 1
AD-mix β	5.05 : 1

^aBased on ¹H NMR



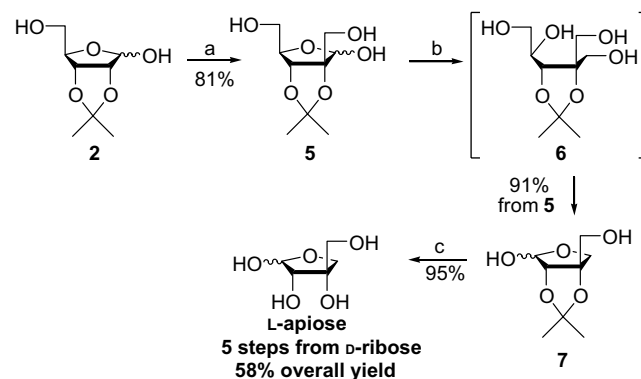
Scheme 2. Synthesis of L-ribose.

the desired stereoselectivity for the L-ribo configuration. First, aldehyde **3** was reacted with osmium tetroxide and *N*-methylmorpholine-*N*-oxide (NMO) to give 2,3-*O*-isopropylidene-L-ribose (**4a**, 56.4% from **1**) and 2,3-*O*-isopropylidene-D-xylose (**4b**, 9.4% from **1**) in 6:1 ratio after the separation by silica gel column chromatography. Stereoselectivity obtained in this reaction is attributed to forming the osmylate in the opposite direction to the bulky 2,3-isopropylidene group. In order to increase the formation of **4a**, asymmetric dihydroxylations using AD-mix α and AD-mix β were employed.

As seen in Scheme 2, stereoselectivity toward **4a** was not improved at all in both cases. Although AD-mix α gave a slightly better result than AD-mix β, both reagents formed **4a** as a major product. Further study on the matching or miss-matching of the catalyst in this dihydroxylation is needed for the explanation of this result. Epoxidation of **3** followed by hydrolysis of the resultant epoxide turned out to be inferior to *cis*-dihydroxylation method in view of yield and stereoselectivity. Finally, compound **4a** was subjected to the hydrolysis using acidic resin to afford L-ribose in 95% yield. L-Ribose was synthesized from D-ribose in five steps and 50% overall yield.

Another L-sugar, L-apiose was also synthesized from D-ribose, as shown in Scheme 3. Aldol condensation¹⁴ of **2** using 35% formaldehyde and potassium carbonate gave 2-*C*-hydroxymethyl derivative **5** as a single stereoisomer in 81% yield. Reduction of **5** followed by oxidative cleavage with sodium metaperiodate afforded the protected L-apiose **7**. Finally, hydrolysis of **7** with acidic resin produced L-apiose in excellent yield.¹² L-Apiose was efficiently synthesized from D-ribose in five steps and 58% overall yield, using stereoselective hydroxymethylation as a key step.

In summary, we have accomplished the efficient and preparative synthesis of L-carbohydrates such as L-ribose and L-apiose, starting from D-ribose. Our synthetic method is regarded as one of the best procedures from the viewpoints of overall yields, number of steps, and



Scheme 3. Synthesis of L-apiose. Reagents and conditions: (a) 35% formaldehyde, K₂CO₃, CH₃OH, reflux, overnight; (b) (i) NaBH₄, CH₃OH, reflux, 1 h; (ii) 0.65 M NaIO₄, CH₂Cl₂, H₂O, 60 °C, 1 h; (c) Dowex-H⁺ resin, 1,4-dioxane-H₂O, 70 °C, 5 h.

mild reaction conditions and will be widely used for the synthesis of biologically active compounds including L-nucleosides.

Acknowledgements

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References and notes

1. Jeong, L. S.; Schinazi, R. F.; Beach, J. W.; Kim, H. O.; Nampalli, S.; Shanmuganathan, K.; Alves, A. J.; McMillan, A.; Chu, C. K. *J. Med. Chem.* **1993**, *36*, 181–195, and references cited therein.
2. Chu, C. K.; Ma, T.; Shanmuganathan, K.; Wang, C.; Xiang, Y.; Pai, S. B.; Tao, G.-Q.; Sommadossi, J.-P.; Cheng, Y.-C. *Antimicrob. Agents Chemother.* **1995**, *39*, 979–981.
3. (a) Lin, T.-S.; Luo, M.-Z.; Liu, M.-C.; Zhu, Y.-L.; Gullen, E.; Dutschman, G. E.; Cheng, Y.-C. *J. Med. Chem.* **1996**, *39*, 1757–1759; (b) Bolon, P. J.; Wang, P.; Chu, C. K.; Gosselin, G.; Boudou, V.; Pierra, C.; Mathe, C.; Imbach, J.-P.; Faraj, A.; el Alaoui, M. A.; Sommadossi, J.-P.; Pai, S. B.; Zhu, Y.-L.; Lin, J.-S.; Cheng, Y.-C.; Schinazi, R. F. *Bioorg. Med. Chem. Lett.* **1996**, *6*, 1657–1662.
4. Lin, T.-S.; Luo, M.-Z.; Liu, M.-C.; Pai, S. B.; Dutschman, G. E.; Cheng, Y.-C. *J. Med. Chem.* **1994**, *37*, 798–803.
5. Holy, A. *Collect. Czech. Chem. Commun.* **1972**, *37*, 4072.
6. (a) Sorbera, L. A.; Castaner, J.; Castaner, R. M.; Bayes, M. *Drugs Future* **2003**, *28*, 870–879; (b) Bryant, M. L.; Bridges, E. G.; Placidi, L.; Faraj, A.; Loi, A. G.; Pierra, C.; Dukhan, D.; Gosselin, G.; Imbach, J. L.; Hernandez, B.; Juodawlkis, A.; Tennant, B.; Korba, B.; Cote, P.; Marion, P.; Cretton-Scott, E.; Schinazi, R. F.; Sommadossi, J. P. *Antimicrob. Agents Chemother.* **2001**, *45*, 229–235.
7. (a) Jung, M. E.; Xu, Y. *Tetrahedron Lett.* **1997**, *38*, 4199–4202; (b) Sivets, G. G.; Klennitskaya, T. V.; Zhernosek, E. V.; Mikhailopulo, I. A. *Synthesis* **2002**, 253–259.
8. Pitsch, S. *Helv. Chim. Acta* **1997**, *80*, 2286–2314.
9. Shi, Z.-D.; Yang, B.-H.; Wu, Y.-L. *Tetrahedron Lett.* **2001**, *42*, 7651–7653.
10. Takahashi, H.; Iwai, Y.; Hitomi, Y.; Ikegami, S. *Org. Lett.* **2002**, *4*, 2401–2403.
11. Nair, V.; Zintek, L. B.; Sells, T. B.; Purdy, D. F.; Jeon, G. S. *Antiviral Res.* **1994**, *23*(Suppl. 1), 38.
12. (a) Koos, M.; Mosher, H. S. *Carbohydr. Res.* **1986**, *146*, 335–341; (b) Ho, P.-T. *Can. J. Chem.* **1979**, *57*, 381–383.
13. Moon, H. R.; Choi, W. J.; Kim, H. O.; Jeong, L. S. *Tetrahedron: Asymmetry* **2002**, *13*, 1189–1190.
14. Ho, P.-T. *Tetrahedron Lett.* **1978**, *19*, 1623–1626.